Enzyme tools and carbohydrate tools for bioproducts development

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GH115 enzymes are the only known enzymes that target GlcA/MeGlcA from high molecular-weight xylans.



In nature, carbonydrate-active enzymes are produced in organisms from various habitats including harsh environmental conditions



http://australianfungi.blogspot.ca



http://wallpapersin4k.



https://en.wikipedia.org/wiki/Iceberg



http://www.bugoutservice.com/termite-

Select bacteria from unique habitats



Marine bacterium Saccharophagus degradans



https://microbewiki.kenyon.edu

Tolerance in high salt condition?



Alkaliphilic bacterium Amphibacillus xylanus



https://microbewiki.kenyon.edu

Tolerance in alkaline condition?

Produce enzymes using biotechnology



towards glucuronoxylans and oligomers with preference towards internally

cubatitutad raciduaa



AxyAgu115A demonstrated better performance in alkaline condition



Consistent with the marine origin of SdeAgu115A, salt activation of enzyme activity was observed for SdeAgu115A but not AxyAgu115A



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AxyAgu115A displayed higher activity towards complex xylans compared to SdeAgu115A

Polymeric substrates	Activity (µmol product/min/µmol enzyme)		
	SdeAgu115A	AxyAgu115A	
Beechwood alucuronoxylan	2470 ± 70	4700 ± 100	
Spruce	917 ± 6	5630 ± 60	
Oat spelt glucuronoarabinoxylan	24 ± 1	501 ± 12	

 Accommodation of complex xylans was consistent with docking analysis that predicted accessibility of AxyAgu115A to branched xylo-oligosaccharides.



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Pros and cons of existing techniques

Separation techniques	Pros	Cons	Re f
Size exclusion	ldentified analytes above 9 kDa	Compounds with the same size co- elute	1
lon chromatogr aphy	Separated neutral and acidic oligomers	Substitution position of MeGlcA was not confirmed	2
HPAEC-PAD	Effective separation of oligomers	Poor compatibility with mass spectrometry due to high salt concentration	3
Capillary electrophor esis	High resolution	Samples typically need derivitation	4

My goal was to develop a LC-MS/MS method that could

- Differentiate diverse characteristics.
- Be label-free.
- Require minimum sample pretreatment.
- Be compatible with mass spectrometry.
- Establish an in-house ms/ms library.
- Be capable of analyzing industrially relevant samples containing a mixture of sugars.
- Be high throughput.

An LC-MS/MS method was developed; over 70 sugars derived from lignocellulose with different sizes, polarity, acidity and linkage positions were identified.

Total ion/mass chromatogram	elative undance tribution	In-hous	e library	Quar asse	ntitative essment
I Mode Precursors NCE Arb	X1 X2	X3 X4	X5 X6	GICA U ^{4m}	² XX XU ^{4m2} XX
9.E+07 X2 X38 109 3000	Sample	[M+COOH]	[M-H ⁺] ⁻	[2M-H ⁺] ⁻	[M + Cl]
	Arabinose ⁿ⁼²	100.00%	$9.50\% \pm 0.3\%$	$0.49\% \pm 0.1\%$	$0.04\% \pm 0.03\%$
Prediction of [27/6H ⁺]	Xvlose ⁿ⁼⁶	100.00%	0.6%	$0.30\% \pm 0.1\%$	$4.00\% \pm 0.5\%$
3.E+07 - M - 3.00	Xylobiose ⁿ⁼⁶	100.00%	$5.79\% \pm 0.1\%$	$2.48\% \pm 0.2\%$	$3.46\% \pm 0.2\%$
Calculate examine ou EF 100 Posicive mode	Xylotriose ⁿ⁼⁶	100.00%	$1.91\% \pm 0.1\%$	$2.83\%\pm0.6\%$	$5.73\% \pm 1.0\%$
mass [21-01-9]=#U7 [M;Na]+	Xylotetraose ⁿ⁼⁶	100.00%	$3.60\% \pm 0.3\%$	NC	$7.50\% \pm 0.8\%$
	Xylopentaose ⁿ⁼⁶	100.00%	$4.86\% \pm 0.3\%$	NC	$8.03\% \pm 0.8\%$
Extract massic 4E-1009 [M+K]+	Xylohexaose ⁿ⁼⁶	100.00%	$9.70\% \pm 0.4\%$	NC	$8.56\% \pm 1.3\%$
	A-X2 ⁿ⁼¹	100.00%	6.1%	6.11%	4.3%
5.E+07	A-X3 ⁿ⁼¹	100.00%	2.0%	NC	11.0%
	Glucuronic Acid	$0.25\% \pm$		$42.50\% \pm$	
	n=2	0.06%	100.00%	40.31%	$0.54\% \pm 0.1\%$
Calculate relative abundance	$\mathbf{U}^{4m2}\mathbf{X}\mathbf{X}^{\mathbf{n}=4}$	ND	100.00%	NC	$\begin{array}{c} 0.15\% \pm 0.1\% \\ 0.10\% \pm \end{array}$
■ 2.E+07 1 1 2.E+07 11 1 11 11 1 11 1 11 1 11 1 11 1 11 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	$\mathbf{X}\mathbf{U}^{4m2}\mathbf{X}\mathbf{X}^{n=2}$	ND	100.00%	NC	0.02%
	0.4 U A	$\mathbf{A}\mathbf{A}_{\mathbf{m}}$ 37.	3 ± 0.1 / 33.22	сот Гм-н]
MS/MS spectra of WM 200 400 600 Concentration (MM) 200 400 600 14 MS/MS spectra of WM 200 400 600 14					

Application of the established LC-MS/MS method to characterize industrial sample before and after enzyme treatment

Compound	Untreated	Enzymes cocktail containing AxyAgu115A
Xylose	++	+++
Xylobiose	++	++
Xylotriose	+++	++++
Xylotetraose	+++	++++
Xylopentaose	+++	++++
Xylohexaose	+++	++++
MeGlcA	+	++++
$\mathrm{U}^{4\mathrm{m}2}\mathrm{X}$	+++	++
$U^{4m2}XX$	+++	++
XU ^{4m2} XX	+++	++

Engineering significance

- SdeAgu115A and AxyAgu115A have the potential to tailor xylans with high molecular weight.
- The unique tolerance properties of these two enzymes can benefit industrial applications with corresponding conditions.
- AxyAgu115A was active on xylan recovered from liquid hot water extraction of mixed hardwood.
- An in-house LC-MS/MS method was developed to characterize the hemicellulosic fraction before and after enzyme treatment, which has the potential to be applied to other plant fibers.

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Fiber characterization

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Synergy

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Application

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Publication Direct and up-close views of plant cell walls show a leading role for ligninmodifying enzymes on ensuing xvlanases





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Biochemical and structural characterization of a five-domain GH115 alpha-glucuronidase from the marine bacterium Saccharophagus degradans 2-40T

Action of a GH115 α-glucuronidase from Amphibacillus xylanus at alkaline condition promotes release of 4-Omethylglucopyranosyluronic acid

subunits of glucuronoxylan and arabinoglucuronoxylan

Manuscript to be submitted Synergistic action of accessory hemicellulases from glycoside hydrolase

families GH115, GH62, and GH51 towards spruce arabinoglucuronoxylan

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LC-MS/MS method development and application of GH115 α -glucuronidase AxyAgu115A, to process glucuronoxylan released through hotwater extraction of mixed hardwood prior to steam-explosion



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